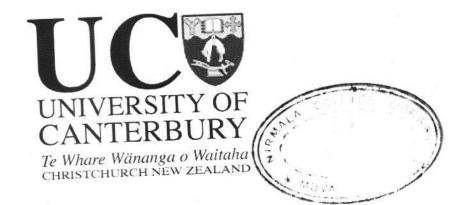
Identification Studies of *Bacillus* Spores Using Fluorescence Spectroscopy

A thesis submitted in partial fulfilment of the requirements for the degree of Doctor of Philosophy in Medical Physics

JOSEPH KUNNIL M.Sc, GDipTch



Department of Physics and Astronomy

University of Canterbury 543.0858 JOS-1 Christ Christ

2005

Supervisor: Associate Professor Lou Reinisch PhD

Abstract

Fluorescence spectroscopy was examined as a potential technique for identifying aerosol particles like bacterial spores. This technique was used for laboratory measurements on some common biological agent simulants. We have measured the intrinsic steady-state fluorescence emission spectra as a function of the excitation wavelength for several bacterial spores (washed and unwashed) in dry and aqueous suspensions at room temperature using excitation wavelengths from 200 to 600 nm. These measurements were compared to those of common, naturally occurring biological components like fungal spores and pollen and non spore samples like ovalbumin. The spectra of samples were combined into fluorescence profiles or fluorescence fingerprints. Different substrates were used for collection and detection of spores. Each bacterium produces a unique in vitro fluorescence profile when measured in dried and aqueous suspension and exhibits a strong maximum in its fluorescence emission spectrum near 330-340 nm. The fluorescence profiles were reproducible. The complexity of microorganisms made the interpretation of their spectral signature a difficult task. Principal components analysis (PCA) and cluster analysis were done as a data reduction technique for detection and identification from different backgrounds. PCA illustrates that linear combination of detected fluorescence intensities, which are present in different ratios in each of samples studied, can be used to discriminate biological agent simulants from other biological samples. The hydration effects, washing effects and the role of tryptophan on spore fluorescence were also investigated. The emission spectra of the dried spores showed a maximum near 330 nm, suggesting a hydrophobic environment for its tryptophan residues. The aqueous solution of tryptophan showed fluorescence shifted to 360 nm and in ethanol solution the maximum was shifted to 340 nm, suggesting a rather more polar average location of the tryptophan. To find the limit of detection we measured the quantum efficiency (QE) of a few samples. We concluded that spectroscopy techniques coupled with effective interpretation models are applicable to biological simulants agents.

Index Heading: Bacteria; Spores; Identification; Fluorescence; Fluorescence Quantum Efficiency; Principal Components Analysis; Cluster Analysis.

Acknowledgments

Many thanks go to Assoc. Prof. Lou Reinisch whose insight and guidance have been invaluable in the struggles of the last few years. The knowledge and the skills, which have been passed on by Prof. Lou, are gratefully acknowledged. Prof. Phil Butler (Head of Department) as co supervisor has also had an influence on these studies. Dr Easaw Chacko of Mathematics and Statistics Department as an associate supervisor has had a leading role in the progress of the research. My special thanks go to two of my friends, Mr. Thomas Francis of Chemistry Department and Mr. Murthy Mittinty of Mathematics and Statistics Department. I also acknowledge my colleagues Ekta Jhala and Sivananthan, S. Also, thankful to the School of Medicine, Christchurch Public Hospital for providing their fluorophotometer for the initial measurement. I gratefully acknowledge the Nova Sol for providing the funding for this research. I also acknowledge the help from the staff of Physical Library especially Mrs Angela Davies. During the course of this work I have been fortunate to have available the wide variety of services provided by all of the technical and office staff of this department. Thanks to the Department of Physics and Astronomy for the financial support throughout this work.

Finally I thank my wife Ann, daughter Vishakham and my son Vanamali for their love and support throughout the study.

l Absti	1 Abstractiii						
2 Ackn	2 Acknowledgments iv						
3 List o	3 List of Figures						
4 List o	of Tablesxv						
1 Intro	duction1						
1.1	1.1 Motivation						
1.2	Bacterial Detection and Identification2						
1.3	Mathematical Models in Identification and Classification of Bacterial						
Spores							
1.4	Overview of the Thesis						
2 Intro	duction to Ultraviolet-Visible Spectroscopy and Data						
Analysis13							
Analys	is13						
Analys	is						
2.1							
2.1 2.1	General Principles of Spectroscopy						
2.1 2.1 2.1	General Principles of Spectroscopy						
2.1 2.1 2.1 2.1	General Principles of Spectroscopy						
2.1 2.1 2.1 2.1 2.1 2.1	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17						
2.1 2.1 2.1 2.1 2.1 2.1	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20						
2.1 2.1 2.1 2.1 2.1 2.1 2.1 2.2	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20.5 Comparison of Fluorescence Quantum Efficiency23						
2.1 2.1 2.1 2.1 2.1 2.1 2.1 2.2	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20.5 Comparison of Fluorescence Quantum Efficiency23Fluorescence Excitation-Emission Spectra via the Two-Dimensional						
2.1 2.1 2.1 2.1 2.1 2.1 2.2 Matri	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20.5 Comparison of Fluorescence Quantum Efficiency23Fluorescence Excitation-Emission Spectra via the Two-Dimensional24						
2.1 2.1 2.1 2.1 2.1 2.1 2.2 Matri 2.3	General Principles of Spectroscopy131 Transmittance and Absorbance152 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20.5 Comparison of Fluorescence Quantum Efficiency23Fluorescence Excitation-Emission Spectra via the Two-Dimensional24K Method24Fluorescence Instrumentation25						
2.1 2.1 2.1 2.1 2.1 2.1 2.2 Matri 2.3 2.4	General Principles of Spectroscopy13.1 Transmittance and Absorbance15.2 Luminescence16.3 The Fluorescence Phenomenon and Frank-Condon Principle17.4 Fluorophores and Biological Fluorescence20.5 Comparison of Fluorescence Quantum Efficiency23Fluorescence Excitation-Emission Spectra via the Two-Dimensional24K Method24Fluorescence Instrumentation25Analysis Data Analysis using Principal Components Analysis and Cluster						

CONTENTS

	3.2	Spectra	al Measurements	33		
	3.3	Spores	Fluorescence	.36		
	3.3.	1 Fluore	escence Quantum Efficiency of Dry Bacillus Spores	36		
	3.3.2 Spores Detection from Tape Substrates/Dusts					
	3.3.	.3 Spores	s Detection from Quartz Substrates/Pollen	42		
	3.3.	.4 Effect	s of Washing on the Identification of Bacillus Spores	43		
	3.3	luorescence of Bacillus Spores and Role of Tryptophan: A				
	Cor	mparativ	e Study of Dry Spores and Spores in Aqueous and Ethanol			
	Sus	spensions	S	. 44		
1	Snoc	troscor	by and Data Analysis Results and Discussions	46		
	Spec	•				
	4.1	Spectra	al Visualization by EEM	. 47		
	4.2	Fluore	scence Quantum Efficiency Measurements of Bacillus Spores	. 52		
	4.2	.1 Absor	bance and Fluorescence of Anthracene in Ethanol	. 52		
	4.2	.2 Quant	tum Efficiency of Bacillus Spores	. 58		
	Z	4.2.2.1	Quantum efficiency of unwashed and washed B. globigii (i) spor	es		
				.59		
	2	4.2.2.2	Quantum efficiency of unwashed and washed B. globigii (ii) Spo	ores		
				.65		
	4	4.2.2.3	Quantum efficiency of unwashed and washed B. globigii (iii)			
	5	Spores		. 69		
	4	4.2.2.4	Quantum efficiency of unwashed B. cereus spores	. 73		
	4.2	.3 Discu	ssions on the Quantum Efficiency Measurements of Bacillus Spor	res		
				. 77		
	4.3	Detect	tion of Bacillus Spores from Different Substrates/Dust/Pollen and			
	Effec	ts of Wa	shing on Fluorescence	. 82		
	4.3	.1 Detec	tion of Spores from Different Substrates	. 82		
	4.3	.2 Detec	tion and Identification of Dry Spores from Dusts Background on			
	Taj	pe Subst	rate	. 84		
	4	4.3.2.1	Spores Fluorescence	. 84		
		4.3.2.2	PCA and Cluster Analysis	. 87		
	4.3	3.3 Detec	ction and Identification of Bacillus Spores from Background Polle	n		
like Pig weed						

			97
4	.3.3.1	Spore Fluorescence	98
4	.3.3.2	PCA and Cluster Analysis	104
4.3	.4 Effec	ets of Washing on Identification of Bacillus Spores	104
	4.3.4.1	Spore Fluorescence	104
	4.3.4.2	PCA and Cluster Analysis	107
		fluorescence of Bacillus spores and Role of Tryptophan: A	
4.4	Autor	e study of Dry Spores and Spores in Aqueous and Ethanol Susper	nsions
Com	parative	e study of Dry Spores and Spores in Addeede and	112
			112
4.4	4.1 Tryp	ptophan Absorption and Fluorescence	115
4.	4.2 Auto	ofluorescence of <i>Bacillus globigii</i> (i)	117
4.	4.3 PCA	A and Cluster Analysis	1 1 /
4.5	Disc	cussions on Spore Fluorescence and Data Analysis	122
4	5 1 Spo	are Fluorescence	122
4.	.5.1 Spo	ta Analysis	131
5 Co	nclusic	ons	135
6 Re	ferenc	es	140
7 AP	PEND	DIX	150
7.1	Lis	st of Publications	158
7.2	2 Ma	athematica Program for Smoothing the Spectrum	160
7.3	3 Ma	athematica Program for Plotting 2D Fluorescence Fingerprints	161
7	1 Pr	incipal Components Model	163
7.	5 Th	he R Program for PCA and Cluster analysis	166
7.	6 P(CA Output	172
1 -	V 1 V		